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Inspiratory muscle training effects on cycling during acute hypoxic exposure

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Corresponding author:

Mitch Lomax, PhD

Department of Sport and Exercise Science

University of Portsmouth

Spinnaker Building

Cambridge Road

Portsmouth

Hampshire

United Kingdom

PO1 2ER

Tel: +44(0)23 9284 5297

Email: [mitch.lomax@port.ac.uk](mailto:mitch.lomax@port.ac.uk)

Heather C Massey, PhD

Department of Sport and Exercise Science

University of Portsmouth

Spinnaker Building

Cambridge Road

Portsmouth

Hampshire

United Kingdom

PO1 2ER

Tel: +44(0)23 9284 3545

Email: [heather.massey@port.ac.uk](mailto:heather.massey@port.ac.uk)

James R House, PhD

Department of Sport and Exercise Science

University of Portsmouth

Spinnaker Building

Cambridge Road

Portsmouth

Hampshire

United Kingdom

PO1 2ER

Tel: +44(0)23 9284 5148

Email: jim.house@port.ac.uk

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## ABSTRACT

*Introduction:* Hypoxic environments increase the physiological demands of exercise. Inspiratory muscle training can reduce the demands of exhaustive exercise in this environment. This study examined the impact of inspiratory muscle training on moderate intensity hypoxic cycling exercise. *Methods:* Seventeen healthy adult men undertook four weeks of inspiratory muscle training (n = 8) or four weeks of sham inspiratory muscle training (n = 9). Subjects completed four fixed- intensity (100 watts) and duration (10 minutes) cycle ergometry tests. Two were undertaken breathing normoxic ambient air, and two breathing a hypoxic gas mixture (14.6% oxygen, balance nitrogen). One normoxic and hypoxic test occurred before, and one after, inspiratory muscle training. *Results:* Inspiratory muscle training increased maximal inspiratory mouth pressure by  $21 \pm 16$  cmH<sub>2</sub>O. Arterial oxygen saturation and its ratio to minute ventilation also increased after inspiratory muscle training during hypoxic exercise from  $83 \pm 4\%$  to  $86 \pm 3\%$  (approximately 3%) and  $2.95 \pm 0.48$  to  $3.52 \pm 0.54$  % $\cdot$ l $\cdot$ min<sup>-1</sup>(approximately 21%), respectively. In addition, minute ventilation, and carbon dioxide output fell by 12-13% after inspiratory muscle training during hypoxic exercise. *Discussion:* Inspiratory muscle training reduced the physiological demand of moderate intensity exercise during acute hypoxic, but not normoxic, exercise. It may therefore be of benefit in adults exercising in a hypoxic environment.

Exercise, altitude, breathing

## INTRODUCTION

Exposure to hypoxia causes a number of cardiopulmonary changes. These include elevated minute ventilation ( $\dot{V}_E$ ), reduced arterial oxygen pressure<sup>24,25</sup> and oxygen saturation<sup>27</sup>, and increased cardiac output ( $\dot{Q}$ )<sup>14,22</sup>.

A hypoxic environment will also affect the body's ability to undertake exercise. For example, a reduction in maximal stroke volume and maximal heart rate<sup>22</sup>, a fall in arterial oxygen content and saturation, diffusion limitation<sup>24,25</sup> and exacerbated hypoxemia<sup>20</sup> will all reduce oxygen delivery to the working muscles during hypoxic exercise. Consequently, maximum oxygen uptake ( $\dot{V}O_2$  max) is lower compared with normoxic exercise<sup>1,2,12</sup>; although submaximal  $\dot{V}O_2$  may<sup>2,14</sup> or may not<sup>14,25</sup> increase. It is therefore not surprising that exercise time to exhaustion is shorter<sup>7,10,23</sup>, the magnitude of peripheral fatigue<sup>1</sup> and dyspnea<sup>7,23</sup> are greater, and diaphragm fatigue is hastened<sup>7,10</sup> during exhaustive hypoxic treadmill (75-85%  $\dot{V}O_2$  max) and cycle ergometry (82-92% peak power output) exercise.

The increase in ventilation that occurs during hypoxic exercise is a compensatory mechanism for the reduced oxygen intake per breath<sup>20</sup>. Consequently,  $\dot{V}_E$  is higher during hypoxic exercise than comparable normoxic exercise<sup>2,23,25</sup>, which increases the oxygen cost of breathing<sup>8</sup>. The breathing musculature will therefore require a larger fraction of oxygen uptake to meet the ventilatory requirements<sup>2,7</sup>. This means that the demands placed upon the lungs and respiratory muscles are greater during hypoxic exercise compared with normoxic exercise<sup>7</sup>. It is therefore not surprising

that inspiratory muscle training (IMT) and respiratory muscle training (RMT) have been employed to reduce the metabolic strain imposed by these muscles in a hypoxic environment.

The breathing muscle training programs adopted in such studies have varied. Some studies have utilized four or eight weeks of respiratory (inspiratory and expiratory) muscle endurance training for five days per week for between 10 and 30 minutes per day<sup>12,16</sup>. Others have utilized four weeks of pressure threshold IMT consisting of only 30 to 40 breaths twice daily, five to seven days per week<sup>10,18</sup>. Regardless of the program adopted, when the fraction of inspired oxygen is reduced to 11% to 14%, IMT has been shown to reduce arterial oxygen desaturation,  $\dot{V}_E$ ,  $\dot{V}O_2$  and  $\dot{Q}$  prior to exhaustion during fixed-intensity (85%  $\dot{V}O_2$  max) hypoxic exhaustive treadmill exercise<sup>10</sup> as well reduce the magnitude of arterial oxygen desaturation at rest<sup>18</sup>. In contrast, RMT has been shown to increase maximal  $V_E$ <sup>12,16</sup> and  $\dot{V}O_2$  max<sup>16</sup> during incremental hypoxic cycling to exhaustion. However, no studies have examined whether or not IMT provides any benefit to non-exhaustive, moderate intensity hypoxic exercise. The aim of the current study was to examine the physiological responses to such exercise. We hypothesised that IMT would reduce the physiological demand of performing moderate intensity non-exhaustive hypoxic exercise.

## METHODS

### Subjects

Seventeen healthy men free from cardiorespiratory disease were randomly assigned to either an IMT (n: 8; age:  $23 \pm 1$  years; body mass:  $80.8 \pm 14.1$  kg; stature:  $1.75 \pm 0.08$  m) or sham IMT (n: 9; age:  $21 \pm 2$  years; body mass:  $78.3 \pm 10.4$  kg; stature:  $1.81 \pm 0.07$  m) group. All subjects were non-smokers. This study protocol was approved in advance by the BioScience Research Ethics Committee, University of Portsmouth. Each subject provided written consent before participating.

#### Equipment and procedures

Subjects completed four fixed- intensity (100 watts) and duration (10 minutes) recumbent cycle ergometry tests in a normobaric chamber (Sporting Edge, Sheffield, UK), two pre (mean  $\pm$  sd: barometric pressure of  $760 \pm 9$  mmHg; temperature of  $21.2 \pm 1.5$  °C; humidity of  $30 \pm 12\%$ ) and two post (mean  $\pm$  sd: barometric pressure of  $765 \pm 7$  mmHg; temperature of  $21.5 \pm 1.1$  °C; humidity of  $41 \pm 12\%$ ) IMT.

Recumbent cycling was chosen for safety reasons and the use of cycling exercise is consistent with past hypoxia and RMT studies<sup>12,16</sup>. Previous work in our laboratory indicated that this intensity could likely be classified as moderate intensity (observed METS ranged from 3.9-4.5) and the duration would be sufficient to reach steady state.

Two tests were undertaken in normoxic conditions (these served as control trials), one pre IMT and one post IMT and two tests while breathing a hypoxic gas mixture (14.6% oxygen, balance nitrogen), one pre IMT and one post IMT. The normobaric chamber used an oxygen filtration system to achieve the pre-set percentage of oxygen, which was confirmed using an independent calibrated hand-held gas analyser (MX6 iBRID, Industrial Scientific, USA). Before the first cycle test, and on

a separate day, subjects attended a pulmonary familiarisation session to practice maximal inspiratory mouth pressure (P<sub>I</sub>max) manoeuvres. These manoeuvres were performed in accordance with BASES guidelines<sup>21</sup>. P<sub>I</sub>max was assessed using a calibrated hand held mouth pressure meter (Micro Medical, Rochester, UK) and was recorded in normoxia pre and post IMT. The coefficient of variation for P<sub>I</sub>max was  $3 \pm 3\%$  and  $8 \pm 9\%$  for the sham IMT and IMT groups respectively.

Before the start of the cycling exercise subjects sat quietly for three minutes. In the normoxic trials subjects breathed the ambient air within the chamber. In the hypoxic trials subjects breathed normoxic air for the first three minutes using an ambient feed from outside of the chamber and then switched to breathing the air within the chamber (14.6% oxygen, balance nitrogen) for a further five minutes after which cycling exercise commenced.

Each cycle test consisted of 10 minutes of fixed intensity seated cycling using an electronically braked ergometer (Lode Angio, Netherlands). The wattage and cadence were fixed throughout corresponding to 100 watts of external work and 70 rpm to 80 rpm, respectively. During this time expired air was collected continuously and passed through a mixing chamber connected to a flow meter (calibrated using a certified 3 litre syringe) and dyspnea was assessed (modified Borg CR10 scale). The percentages of expired oxygen and carbon dioxide were determined using a gas analyser (G1R250 gas analyser, Hitech Instruments, UK), which was calibrated using ambient normoxic air fed through a sample tube, and gases of a known concentration. Expired  $\dot{V}_E$ ,  $\dot{V}O_2$ , carbon dioxide output ( $\dot{V}CO_2$ ), respiratory

exchange ratio (RER), tidal volume (VT) and breathing frequency ( $f_r$ ) were recorded continuously (Power Lab SP16 analogue to digital converter, AD Instruments, Australia) and displayed on a portable computer using data acquisition software (Chart 6, AD Instruments, Australia). The time delay inherent in measuring expired oxygen and carbon dioxide fractions with this method was rectified prior to data analysis.

Heart rate ( $f_c$ ) and arterial oxygen saturation were measured continuously from a 3-lead ECG (Life Pulse, UK) and pulse oximetry ( $SpO_2$ ) (NONIN, 7500, USA), respectively. Data were recorded using data acquisition software (Chart 6, AD Instruments, Australia) for subsequent analysis. The ratio of  $SpO_2/\dot{V}_E$ , which provides an estimate of ventilatory efficiency<sup>3</sup>, was calculated after each test.

Excluding the P<sub>I</sub>max and dyspnea data, all remaining data are collectively referred to as metabolic data.

After completion of the normoxic and hypoxic pre IMT trials, subjects completed four weeks of IMT or sham IMT using a commercially available inspiratory muscle trainer (POWERbreathe®, HaB International, Warwickshire, UK). Subjects completed two sets of 30 breaths daily (one set in the morning and one set in the evening), seven days a week in a seated position. In the IMT group, the load on the inspiratory muscle trainer was set-up in the laboratory to reflect 50% of each subject's highest baseline P<sub>I</sub>max. Conversely, the sham IMT group breathed through the trainer but against no load. Subjects in the IMT group were instructed to increase the load weekly so that they could only just complete 30 breaths<sup>18</sup>. All subjects kept a weekly IMT training log for subsequent evaluation of IMT adherence ( $88 \pm 10\%$

and  $77 \pm 0.1\%$  of breath repetitions completed in the IMT and sham UMT groups respectively). The two remaining normoxic and hypoxic trials were completed within five days of completing IMT.

#### Statistical analysis

Data were assessed for normality using the Shapiro-Wilk test, skewness and kurtosis. All breath-by-breath data,  $SpO_2$ ,  $f_c$  and  $SpO_2/\dot{V}_E$ , and dyspnea were averaged per minute with only the second half of each test analysed<sup>14,24</sup>. Two-way factorial analysis of variance (ANOVA) were used to assess for differences between groups (IMT vs. sham), simulated altitude (normoxia vs. hypoxia) and IMT status (pre vs. post IMT) per dependent variable. In the case of P<sub>I</sub>max, a one-way factorial ANOVA was used to assess for differences between groups (IMT vs. sham) and IMT status (pre vs. post IMT): the highest baseline value recorded pre IMT and post IMT was used regardless of simulated altitude trial. Delta change in P<sub>I</sub>max pre and post cycle ergometry was used to assess for inspiratory muscle fatigue (IMF) pre and post IMT per simulated altitude trial and was also assessed using repeated measures factorial ANOVA.

Post hoc analyses were undertaken using one-way repeated measures ANOVAs, paired samples t-tests and independent samples t-tests, where relevant and the p value adjusted accordingly.

Where relevant, effect sizes were calculated using Cohen's *d* with an effect size of 0.2 deemed small, 0.5 medium and above 0.8 deemed large<sup>9</sup>. *P* was set at 0.05 and

analyses were undertaken using IBM SPSS statistics version 22. Unless otherwise stated data are presented as mean  $\pm$  sd.

## RESULTS

An interaction was observed between IMT status and group for P<sub>I</sub>max ( $F_1 = 8.502$ ,  $P = 0.011$ ). P<sub>I</sub>max was higher in the IMT group both before (IMT group:  $155 \pm 17$  cmH<sub>2</sub>O; sham group:  $123 \pm 26$  cmH<sub>2</sub>O;  $P = 0.010$ ) and after (IMT group:  $176 \pm 27$  cmH<sub>2</sub>O; sham group:  $125 \pm 18$ ;  $P < 0.001$ ) IMT. Importantly however, sham IMT had no effect on P<sub>I</sub>max ( $P = 0.715$ ) but P<sub>I</sub>max was increased by  $21 \pm 16$  cmH<sub>2</sub>O in the IMT group following IMT ( $P = 0.008$ ,  $d = -0.93$ ). Thus, we can be confident that IMT did increase P<sub>I</sub>max in the IMT group but not the sham group. Additionally, IMF was not observed following any trial.

It was unsurprising that VT ( $F_1 = 6.264$ ,  $P = 0.024$ ),  $f_r$  ( $F_1 = 11.082$ ,  $P = 0.005$ ),  $f_c$  ( $F_1 = 13.525$ ,  $P = 0.005$ ),  $\dot{V}_E$  ( $F_1 = 33.484$ ,  $P < 0.001$ ),  $\dot{V}O_2$  ( $F_1 = 7.154$ ,  $P = 0.018$ ), RER ( $F_1 = 21.820$ ,  $P < 0.001$ ) and dyspnea ( $F_1 = 29.562$ ,  $P < 0.001$ ) were all higher in hypoxic exercise compared with normoxic exercise, regardless of IMT status or group. Similarly, SpO<sub>2</sub> ( $F_1 = 201.130$ ,  $P < 0.001$ ) and SpO<sub>2</sub>/ $\dot{V}_E$  ( $F_1 = 137.384$ ,  $P < 0.001$ ) were also lower during hypoxic exercise (Tables I and II).

\*\*Tables I and II about here\*\*

Importantly, IMT but not sham IMT was associated with a reduction in  $\dot{V}_E$  ( $F_1 = 15.484$ ,  $P = 0.001$ ) and  $\dot{V}CO_2$  ( $F_1 = 5.570$ ,  $P = 0.042$ ) during hypoxic, but not normoxic exercise. In contrast,  $SpO_2$  ( $F_1 = 29.525$ ,  $P = 0.004$ ) and  $SpO_2/\dot{V}_E$  ( $F_1 = 10.781$ ,  $P = 0.005$ ) were increased following IMT in the IMT group during hypoxic exercise (Figure 1). Interestingly, dyspnea fell following both hypoxic and normoxic exercise in response to both IMT and sham IMT ( $F_1 = 18.255$ ,  $P = 0.001$ ) but no difference was observed in dyspnea between the groups.

\*\*\*Figure 1 about here\*\*\*

## DISCUSSION

The aim of this study was to determine if IMT could lower the physiological demands of moderate intensity non-exhaustive cycling exercise in a hypoxic atmosphere (14.6% oxygen), which simulated an altitude of approximately 3000 m. Our data indicate that four weeks of IMT increased  $P_{Imax}$  and reduced  $\dot{V}_E$ ,  $\dot{V}CO_2$ , the magnitude of arterial oxygen de-saturation, and improved ventilatory efficiency, in response to an acute hypoxic exposure during moderate intensity fixed-rate cycling exercise. This is the first study to demonstrate that exercise does not need to be exhaustive for IMT to be of benefit when exercising at a simulated high altitude. Furthermore, as there was no evidence of IMF our data also suggest that IMT can be beneficial in hypoxia even in the absence of such fatigue.

As expected,  $\dot{V}_E$ ,  $\dot{V}O_2$ ,  $fc$  and dyspnea were greater, and  $SpO_2$  lower, during hypoxic exercise compared with normoxic exercise (Tables I and II). Interestingly, studies

using exhaustive high-intensity cycling ergometry (82-92% of maximal work load and 75% of  $\dot{V}O_2$  max) have shown that the increase in  $\dot{V}_E$  can be mediated via  $f_r$ <sup>1,7</sup> or  $VT$ <sup>23</sup> when breathing a hypoxic gas mixture of 9% to 15% oxygen. We observed an increase in both  $f_r$  and  $VT$  with no consistent pattern evident between tests.

Of greater relevance to this study however, was the impact of IMT on hypoxic exercise. Following IMT  $\dot{V}_E$  was reduced in the hypoxic trial but only in the IMT group (Figure 1). The individual effects of  $VT$  and  $f_r$  following IMT were not significant and only in combination were they able to impact  $\dot{V}_E$ . Nevertheless, the effect size data suggest that the reduction was predominantly mediated via a fall in  $VT$  (medium to large effect size) rather than  $f_r$  (small effect size) (Table II). This is consistent with the findings of Downey et al.<sup>10</sup>

We do not believe that this fall in  $\dot{V}_E$  was due to a reduction in the work of breathing *per se*. Two key observations support this. Firstly, as absolute workload and duration were fixed (100 watts and 10 minutes, respectively), a fall in the work of breathing would be evident during both normoxic and hypoxic exercise. However,  $\dot{V}_E$  only declined during hypoxic exercise (Table II). Secondly, had the work of breathing declined post IMT, the oxygen requirement of the inspiratory muscles would have fallen and  $\dot{V}O_2$  would have been lower during both normoxic and hypoxic exercise<sup>15</sup>. However,  $\dot{V}O_2$  demonstrated a non-significant increase during normoxic exercise (small to medium effect size) and a non-significant fall (large effect size) during hypoxic exercise (Table II). It is more likely that the fall in  $\dot{V}_E$  in hypoxia post IMT was linked to the 3% increase in  $SpO_2$ . As a result, the  $SpO_2/\dot{V}_E$

ratio increased (Figure 1) indicating that the amount of ventilation required to achieve a given level of oxygen saturation fell<sup>3</sup>.

The question of how IMT offers protection against the magnitude of fall in arterial oxygen saturation and in-turn reduces  $\dot{V}_E$  is yet to be fully elucidated. An increase in lung diffusion capacity is a potential mechanism<sup>10</sup> and could also explain the fall in  $\dot{V}CO_2$ , the tendency for  $\dot{V}O_2$  to fall (*albeit* non-significantly) and the increase in ventilatory efficiency ( $SpO_2/\dot{V}_E$  ratio) observed during hypoxic exercise following IMT in the current study (Figure 1). Indeed, diffusion limitation (and ventilation-perfusion mismatch) have been observed during light intensity ( $\dot{V}O_2$  above 1.0 l·min<sup>-1</sup>) steady-rate cycling exercise of 7-9 minutes at a simulated altitude of 3,048m<sup>24</sup>, which is not dissimilar to the conditions of the current study. Importantly, Downey et al.<sup>10</sup> have found that lung diffusion capacity increased by 23% during hypoxic exercise following four weeks of IMT and  $\dot{Q}$  and  $\dot{V}_E$  decreased by 13% and 25%, respectively. They suggested that the time available for gas exchange to occur in the lung was likely increased as a result of pulmonary diffusion and  $\dot{Q}$  changes. This in-turn would lengthen the red blood cell transit time promoting oxygen-haemoglobin binding and hence increase  $SpO_2$ <sup>10</sup>. Downey et al.<sup>10</sup> postulate that the increase in  $SpO_2$  might reduce the peripheral chemoreceptor input and in-turn lower  $\dot{V}_E$ .

It should be noted that the metabolic responses observed in the current study and by Downey et al.<sup>10</sup> are different to those reported at exhaustion following an incremental cycle exercise test in hypoxic (11%-12% inspired oxygen) post RMT. Both Keramidas et al.<sup>16</sup> and Esposito et al.<sup>12</sup> reported an increase in maximum  $\dot{V}_E$  (12%) at

the point of exhaustion, while Keramidas et al.<sup>16</sup> also observed an increase in  $\dot{V}O_2$  max, although cycle time to exhaustion was unaffected. Thus, it appears that IMT is capable of reducing  $\dot{V}_E$  and  $\dot{V}CO_2$  during submaximal cycling (present study) or running exercise prior to the point of exhaustion<sup>10</sup>, whereas RMT can increase maximal  $\dot{V}_E$  during incremental exhaustive cycling exercise and possibly even  $\dot{V}O_2$  max depending on the level of ventilation-perfusion mismatch<sup>12,16</sup>.

Given the complexity of the pulmonary system, and the different exercise and IMT/RMT protocols adopted in past hypoxic studies, delineating exactly how IMT benefits exercise in hypoxia is challenging. It is important to state that the ergogenic effect of IMT is unlikely to be due simply to an increase in inspiratory muscle strength alone<sup>13</sup>. While we are not aware of any studies showing that IMT can increase lung growth, an increase in lung volume has been observed following IMT<sup>11</sup> and it has been demonstrated that changes in the perceptual and cardiovascular responses to normoxic exercise following IMT are secondary to IMT-induced structural and functional changes within the inspiratory muscles<sup>13</sup>. For example, it has been shown that following IMT the inspiratory muscle metaboreflex is attenuated during plantarflexion exercise<sup>19</sup>,  $f_c$  and the perceptions of breathing and leg effort are reduced during treadmill walking while carrying a 25 kg back pack<sup>13</sup>, and blood lactate concentration is reduced during hyperpnea<sup>4</sup>. Clearly, more studies are needed to delineate how the morphological changes arising from IMT contribute to enhanced exercise performance. Moreover, as the mechanics of breathing are altered in hypobaric hypoxia because air density is lower and pulmonary blood flow is increased<sup>6</sup>, careful consideration should be given as to whether hypobaric or normobaric hypoxic experimental conditions are adopted.

Furthermore, our SpO<sub>2</sub> data raise an interesting question regarding the potential role of IMT in reducing acute mountain sickness (AMS). The occurrence of AMS following acute exposure to altitude (i.e. minutes to hours) is linked with low SpO<sub>2</sub> values<sup>5,17</sup>. Loeppky et al.<sup>17</sup> reported that sufferers of early-onset (within 8 to 12 hours of hypoxic exposure) AMS exhibited greater hypoxemia than non-sufferers despite an equivalent ventilatory response. The fall in SpO<sub>2</sub> was therefore not because of a lower ventilatory response which can reduce SpO<sub>2</sub><sup>5</sup>. Although our data provide no evidence that IMT can offer protection against AMS on initial exposure to altitude, it does provide a rationale for investigating this possibility.

Lastly, it would be remiss to ignore our dyspnea data and the baseline differences in P<sub>I</sub>max between the sham IMT and IMT group. That dyspnea fell during hypoxic exercise post IMT during hypoxic exercise in both the sham IMT and IMT groups should be acknowledged, if only to indicate that IMT was unable to modify dyspnea at the exercise intensity and level of hypoxia (14.6% oxygen) adopted. Thus, the fall in dyspnea that we observed might simply reflect a perceptual familiarisation arising from a second exposure to hypoxia.

The difference in baseline P<sub>I</sub>max between groups occurred because we randomly allocated subjects to one of the groups instead of matching subjects based on inspiratory muscle strength. While we acknowledge this as a limitation and recommend that future studies should endeavour to match for P<sub>I</sub>max, we do not believe that this invalidates our findings as no significant differences were observed between groups in  $\dot{V}_E$ ,  $\dot{V}_{O_2}$ ,  $\dot{V}_{CO_2}$ , VT,  $f_r$  and SpO<sub>2</sub> pre IMT. Furthermore, P<sub>I</sub>max only increased following IMT (average increase of 21 cmH<sub>2</sub>O) not sham IMT

(average increase of 1 cmH<sub>2</sub>O), indicating that a training effect did take place.

Rather, such a discrepancy in baseline inspiratory muscle strength would be more likely to curtail the potential gain for an improvement.

In conclusion, four weeks of pressure-threshold IMT exerted beneficial physiological effects during moderate intensity ( $\dot{V}_E \leq 30 \text{ l}\cdot\text{min}^{-1}$  and  $\dot{V}O_2 < 1.28 \text{ l}\cdot\text{min}^{-1}$ ) constant-load (100 watts), non-exhaustive, fixed-duration (10 minutes) cycling exercise in acute hypoxia (14.6% oxygen). In contrast, no such benefits were observed post IMT during identical exercise in normoxia. Specifically,  $\dot{V}_E$  and  $\dot{V}CO_2$  were lower, and  $SpO_2$  and  $SpO_2/\dot{V}_E$  higher, during hypoxic exercise after IMT. Interestingly, these changes occurred in the absence of inspiratory muscle fatigue, indicating that such fatigue does not need to be present for IMT to be of benefit. Given that IMT was able to modify the initial  $\dot{V}_E$  and  $SpO_2$  response in acute hypoxia, it should be investigated whether or not IMT can reduce the likelihood or severity of AMS.

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## REFERENCES

1. Amann M, Pegelow DF, Jacques AJ, Dempsey JA. Inspiratory muscle work in acute hypoxia influences locomotor muscle fatigue and exercise performance of healthy humans. *Am J Physiol.* 2007;293(5):R2036-R2045.
2. Benoit H, Busso T, Prieur F, Castells J, Freyssenet D, Lacour JR, Denis C, Geysant A. Oxygen uptake during submaximal incremental and constant work load exercise in hypoxia. *Int J Sports Med.* 1997;18(2):101-105.
3. Bernardi L, Schneider A, Pomidori, Paolucci E, Cogo A. (2006). Hypoxic ventilatory response in successful extreme altitude climbers. *Eur Respir J.* 2006;27(1):165-171.
4. Brown PI, Sharpe GR, Johnson MA. Inspiratory muscle training reduces blood lactate concentration during volitional hyperpnoea. *Eur J Appl Physiol.* 2008;104(1):111-117.
5. Burtcher M, Flatz M, Faulhaber M. Prediction of susceptibility to acute mountain sickness by SaO<sub>2</sub> values during short-term exposure to hypoxia. *High Alt Med Biol.* 2004;5(3):335-340.
6. Congo A, Legnani D, Allegra L. Respiratory function at different altitudes. *Respiration.* 1997;64:416-421.
7. Cibella F, Cuttitta G, Kayser B, Narici M, Romano S, Saibene F. Respiratory mechanics during exhaustive submaximal exercise at high altitude in healthy humans. *J Physiol.* 1996;494(3):881-890.

8. Coast JR, Rasmussen SA, Krause KM, O’Kroy JA, Loy RA, Rhodes J. Ventilatory work and oxygen consumption during exercise and hyperventilation. *Journal of Applied Physiology*. 1993;74(2):793-798.
9. Cohen J. *Statistical power analysis for the behavioural sciences*. Hillsdale NJ: Lawrence Erlbaum Associates;1988.
10. Downey AE, Chenoweth LM, Townsend DK, Ranum JD, Ferguson CS, Harms CA. Effects of inspiratory muscle training on exercise responses in normoxia and hypoxia. *Respir Physiol Neurobiol*. 2007;156(2):137-146.
11. Enright SJ, Unnithan VB, Heward C, Withnall L, Davies DH. Effect of high-intensity inspiratory muscle training on lung volumes, diaphragm thickness, and exercise capacity in subjects who are healthy. *Phys Ther*. 2006;86(3):345-354.
12. Esposito F, Limonta E, Alberti G, Veicsteinas A, Ferretti G. Effect of respiratory muscle training on maximum aerobic power in normoxia and hypoxia. *Respir Physiol Neurobiol*. 2010;170(3):268-272.
13. Faghy MA, Brown PI. Training the inspiratory muscles improves running performance when carrying a 25 kg thoracic load in a backpack. *Eur J Sports Sci*. 2015;16(5):585-594.
14. Gale GE, Torre-Bueno JR, Moon RE, Saltzman HA, Wagner PD. Ventilation-perfusion inequality in normal humans during exercise at sea level and simulated altitude. *J Appl Physiol*. 1985;58(3):978-988.
15. Harms CA, Wetter TJ, McClaren SR, Pegelow DF, Nickle GA, Nelson W, Hanson P, Dempsey JA. Effects of respiratory muscle work on cardiac output and its distribution during maximal exercise. *J Appl Physiol*. 1998;85(2):609-618.

16. Keramidis ME, Debevec T, Amon M, Kounalakis SN, Simunic B, Mekjavic IB. Respiratory muscle endurance training: effect on normoxic and hypoxic exercise performance. *Eur J Appl Physiol.* 2010;108(4):759-769.
17. Loeppky JA, Icenogle MV, Charlton GA, Conn CA, Maes D, Riboni K, Gates L, Melo MF, Roach RC. Hypoxemia and acute mountain sickness: which comes first? *High Alt Med Biol.* 2008;9(4):271-279.
18. Lomax M. Inspiratory muscle training, altitude, and arterial oxygen desaturation: a preliminary investigation. *ASEM.* 2010;81(5):498-501.
19. McConnell AK, Lomax M. The influence of inspiratory muscle work history and specific inspiratory muscle training upon human limb muscle fatigue. *J Physiol.* 2006;557(1):445-457.
20. Marconi C, Cerretelli P. Altitude physiology: the impact of hypoxia on human performance. In: Taylor NAS, Graeller H, editors. *Physiological bases of human performance during exercise.* Edinburgh: Churchill Livingstone; 2008. p. 433-446.
21. McConnell AK. Lung and respiratory muscle function. In: Winter EM, Jones AM, Davison RCR, Bromely PD, Mercer TH, editors. *Sport and exercise physiology testing guidelines (BASES) vol 2: exercise and clinical testing.* UK: Routledge; 2007. p. 63-75.
22. Naeije R. Physiological adaptation of the cardiovascular system to high altitude. *Prog Cardiovasc Dis.* 2010;52:456-466.
23. Romer LM, Haverkamp HC, Amann M, Lovering AT, Pegelow DF, Dempsey JA. Effect of acute severe hypoxia on peripheral fatigue and endurance capacity in healthy humans. *Am J Physiol.* 2007;292(1):R598-R606.

24. Torre-Bueno JR, Wagner PD, Saltzman HA, Gale GE, Moon RE. Diffusion limitation in normal humans during exercise at sea level and simulated altitude. *J Appl Physiol.* 1985;58(3):989-995.

25. Wagner PD, Gale GE, Moon RE, Torre-Bueno JR, Stolp BW, Saltzman HA. Pulmonary gas exchange in humans exercising at sea level and simulated altitude. *J Appl Physiol.* 1986;61(1):260-270.

26. Wilkins BE, Schrage WG, Liu Z, Hancock KC, Joyner MJ. Systematic hypoxia and vasoconstrictor responsiveness in exercising human muscle. *J Appl Physiol.* 2006;101(5):1343-1350.

Table I. Pre and post IMT data averaged over the second half of 10 minutes fixed-intensity normoxic cycle ergometry: mean  $\pm$  sd

Measure	IMT group			Sham IMT		
	Pre IMT	Post IMT	<i>d</i>	Pre IMT	Post IMT	<i>d</i>
$\dot{V}_E$ (l·min <sup>-1</sup> )	22. $\pm$ 3.4 <sup>§§</sup>	24.1 $\pm$ 3.1	-0.49	22.7 $\pm$ 1.4 <sup>§§</sup>	22.5 $\pm$ 3.6 <sup>§§</sup>	0.07
$\dot{V}O_2$ (l·min <sup>-1</sup> )	1.16 $\pm$ 0.14	1.22 $\pm$ 0.13	-0.44	1.21 $\pm$ 0.11	1.19 $\pm$ 0.24	0.11
$\dot{V}CO_2$ (l·min <sup>-1</sup> )	1.05 $\pm$ 0.08	1.04 $\pm$ 0.11	-0.10	1.08 $\pm$ 0.08	1.03 $\pm$ 0.11	0.09
RER	0.92 $\pm$ 0.06	0.87 $\pm$ 0.10 <sup>§§</sup>	0.61	0.90 $\pm$ 0.07	0.89 $\pm$ 0.13	0.10
VT (l)	1.30 $\pm$ 0.20 <sup>§</sup>	1.36 $\pm$ 0.20	-0.30	1.17 $\pm$ 0.08	1.12 $\pm$ 0.21	0.31
$f_r$ (breaths·min <sup>-1</sup> )	21 $\pm$ 5	21 $\pm$ 5	0.00	23 $\pm$ 2 <sup>§</sup>	22 $\pm$ 4	0.17
SpO <sub>2</sub> (%)	96 $\pm$ 2 <sup>§§</sup>	96 $\pm$ 1 <sup>§§</sup>	0.00	96 $\pm$ 1 <sup>§§</sup>	95 $\pm$ 1 <sup>§§</sup>	1.00
SpO <sub>2</sub> / $\dot{V}_E$ (%·l·min <sup>-1</sup> )	4.37 $\pm$ 0.80 <sup>§§</sup>	402 $\pm$ 0.58 <sup>§</sup>	0.50	4.31 $\pm$ 0.27 <sup>§§</sup>	4.38 $\pm$ 0.67 <sup>§§</sup>	-0.14
$f_c$ (b·min <sup>-1</sup> )	129 $\pm$ 17	132 $\pm$ 16 <sup>*§</sup>	-0.18	125 $\pm$ 7	112 $\pm$ 11 <sup>**§§</sup>	1.41
Dyspnea	2.3 $\pm$ 1.5 <sup>§§</sup>	2.0 $\pm$ 0.8 <sup>§</sup>	0.25	2.5 $\pm$ 0.4 <sup>§§</sup>	2.4 $\pm$ 0.7	0.18

Notes. \*( $P \leq 0.05$ ) \*\*( $P \leq 0.01$ ) different to pre IMT within-trial (pre, post IMT). §( $P \leq 0.05$ )  
§§( $P \leq 0.01$ ) different to hypoxia per group (sham, IMT). *d* = Cohen's *d*.

Table II. Pre and post IMT data averaged over the second half of 10 minutes fixed-intensity hypoxic cycle ergometry: mean  $\pm$  sd

Measure	IMT group			Sham IMT		
	Pre IMT	Post IMT	<i>d</i>	Pre IMT	Post IMT	<i>d</i>
$\dot{V}_E$ (l·min <sup>-1</sup> )	28.9 $\pm$ 5.1	24.9 $\pm$ 4.3*	0.85	24.9 $\pm$ 2.1	26.9 $\pm$ 3.9	-0.64
$\dot{V}O_2$ (l·min <sup>-1</sup> )	1.10 $\pm$ 0.15	0.97 $\pm$ 0.14	0.90	1.06 $\pm$ 0.14	1.27 $\pm$ 0.20	-1.27
$\dot{V}CO_2$ (l·min <sup>-1</sup> )	1.14 $\pm$ 0.18	1.00 $\pm$ 0.16*	0.82	1.10 $\pm$ 0.09	1.12 $\pm$ 0.10	-0.21
RER	1.06 $\pm$ 0.14	1.03 $\pm$ 0.11	0.25	1.05 $\pm$ 0.13	0.92 $\pm$ 0.10	1.12
VT (l)	1.57 $\pm$ 0.33	1.34 $\pm$ 0.31	0.72	1.21 $\pm$ 0.13	1.27 $\pm$ 0.11	-0.50
$f_r$ (breaths·min <sup>-1</sup> )	23 $\pm$ 7	22 $\pm$ 5	0.16	24 $\pm$ 3	25 $\pm$ 5	-0.24
SpO <sub>2</sub> (%)	83 $\pm$ 4	86 $\pm$ 3*	-0.85	85 $\pm$ 3	82 $\pm$ 5*	0.73
SpO <sub>2</sub> / $\dot{V}_E$ (%·l·min <sup>-1</sup> )	2.95 $\pm$ 0.48	3.52 $\pm$ 0.54*‡	-1.15	3.62 $\pm$ 0.66	3.08 $\pm$ 0.42	0.98
$f_c$ (b·min <sup>-1</sup> )	138 $\pm$ 14	135 $\pm$ 18	0.19	129 $\pm$ 12	123 $\pm$ 12	0.50
Dyspnea	4.8 $\pm$ 1.8	2.8 $\pm$ 1.0**	1.37	3.8 $\pm$ 1.1	2.9 $\pm$ 1.0*	0.86

Notes. \*( $P \leq 0.05$ ) \*\*( $P \leq 0.01$ ) different to pre IMT within-trial (pre, post IMT). ‡( $P \leq 0.05$ ) different to sham. *d* = Cohen's *d*.

Figure 1. Post intervention sham IMT group (closed bars) and IMT group (open bars) hypoxic data expressed as percentage change from pre IMT data: mean  $\pm$  sd

Notes.  $*(P < 0.05)$  different to within-trial for the IMT group only.  $^{\ddagger}(P < 0.05)$   $^{\ddagger\ddagger}(P < 0.01)$  different to sham IMT group post IMT.